

STRUCTURE AND SYNTHESIS OF UNUSUAL CYTOKININ METABOLITES

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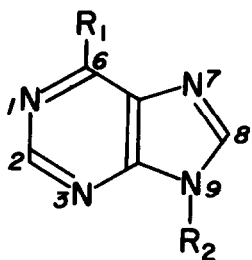
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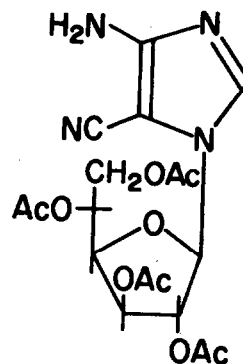
(Received in UK 27 January 1975; accepted for publication 12 February 1975)

There have been several recent reports¹⁻⁵ of the isolation from a range of plant tissues of 7- and 9-glucosides as stable metabolites of the cytokinins, zeatin and 6-benzylaminopurine (6-BAP). From an analysis of the mass spectrum of the trimethylsilyl (TMS) derivative of the 7-glucosyl metabolite of 6-BAP, Fox¹ suggested that the sugar moiety possessed the furanose structure. On the other hand, it has been shown that 9-glucosylzeatin, the principal metabolite of zeatin in Zea mays seedlings, and 6-benzylamino-9-glucosylpurine, a major metabolite of 6-BAP in de-rooted radish seedlings, could not be differentiated from the synthetic 9- β -D-glucopyranosyl derivatives of zeatin (1) and 6-BAP (3) respectively³. The minute quantities (<100 μ g) of isolated metabolites restricted measurements to u.v., mass spectral and t.l.c. comparisons.

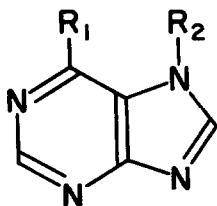
We have synthesised the 9- β -D-glucopyranosides of zeatin (2) and 6-BAP (4) by standard methods^{6,7,8} and compared their u.v. and mass spectral, g.c.-m.s. (permethyl and TMS derivatives) and t.l.c. (with and without borate) characteristics with those of the synthetic³ 9- β -D-glucopyranosides (1) and (3) and the 9-glucosyl metabolites of zeatin and 6-BAP respectively. The results confirmed that the metabolites were 9-glucopyranosides. As the 9-substituted metabolites were not hydrolysed by either α - or β -glucosidase, the nature of the anomeric linkage in the 9-glucosyl metabolite of 6-BAP was established as β using the above criteria by its identity with the synthetic 9- β -D-glucopyranoside (3) and dissimilarity to the synthetic 9- α -D-glucopyranoside of 6-BAP (5). The latter compound (5) was prepared by fusion of the triphenylsilyl derivative of 6-chloropurine with α -D-tetra-O-acetylglucopyranosyl-bromide, separation of the 9- α from the 9- β -anomer (formed in the ratio 2:3) and subsequent



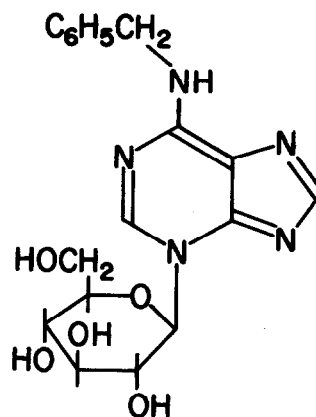
- (1) $R_1 = \begin{array}{c} \text{HOCH}_2 \\ | \\ \text{C}=\text{C} \\ / \quad \backslash \\ \text{CH}_3 \quad \text{H} \\ \quad \quad \quad | \\ \quad \quad \quad \text{CH}_2\text{NH}- \end{array}$; $R_2 = \beta\text{-D-glucopyranosyl-}$
 (2) $R_1 = \quad \quad \quad "$; $R_2 = \beta\text{-D-glucofuranosyl-}$
 (3) $R_1 = \text{C}_6\text{H}_5\text{CH}_2\text{NH-}$; $R_2 = \beta\text{-D-glucopyranosyl-}$
 (4) $R_1 = \quad \quad \quad "$; $R_2 = \beta\text{-D-glucofuranosyl-}$
 (5) $R_1 = \quad \quad \quad "$; $R_2 = \alpha\text{-D-glucopyranosyl-}$



(11)



- (6) $R_1 = \begin{array}{c} \text{HOCH}_2 \\ | \\ \text{C}=\text{C} \\ / \quad \backslash \\ \text{CH}_3 \quad \text{H} \\ \quad \quad \quad | \\ \quad \quad \quad \text{CH}_2\text{NH}- \end{array}$; $R_2 = \beta\text{-D-glucopyranosyl-}$
 (7) $R_1 = \quad \quad \quad "$; $R_2 = \beta\text{-D-glucofuranosyl-}$
 (8) $R_1 = \text{C}_6\text{H}_5\text{CH}_2\text{NH-}$; $R_2 = \beta\text{-D-glucopyranosyl-}$
 (9) $R_1 = \quad \quad \quad "$; $R_2 = \beta\text{-D-glucofuranosyl-}$
 (10) $R_1 = \text{CH}_3\text{S-}$; $R_2 = \beta\text{-D-glucofuranosyl-}$



(12)

displacement of the 6-chloro-substituent and concomitant de-acetylation with benzylamine in refluxing *n*-butanol. A mixture of the α - and β -anomers (5) and (3) in the ratio (1:2) was also obtained in good yield on condensing α -D-tetra-0-acetylglucopyranosylbromide with the 9-tricarbonylcyclohexa-2,4-dienyliron derivative of 6-BAP (obtained by treating 6-BAP with $[\text{C}_6\text{H}_7\text{Fe}(\text{CO})_3]\text{BF}_4$) in DMF at 90° followed by treatment with methanolic ammonia.

7- β -D-Glucofuranosyl-6-methylthiopurine (10) was prepared via ring closure of 4-amino-5-cyano-1-tetra-0-acetyl- β -D-glucofuranosylimidazole (11) using the procedure of Townsend *et al.*,⁹ substituting penta-0-acetylglucofuranose¹⁰ for tetra-0-acetylribofuranose in the first step. Displacement of the 6-methylthio substituent in (10) using neat benzylamine at 140° for 24 hours gave 6-benzylamino-7- β -D-glucofuranosylpurine (9) while displacement with *trans*-4-amino-2-methylbut-2-enol in refluxing *n*-butanol afforded 7- β -D-glucofuranosylzeatin (7). Both cytokinin glucofuranosides (7) and (9) were also synthesised in better yield from the ethoxymethyleneimine of the above imidazole (11) and utilising benzylamine or *trans*-4-amino-2-methylbut-2-enol to effect pyrimidine ring closure¹¹ to the 1-substituted adenine derivatives. A modified Dimroth rearrangement¹² of these intermediates provided the desired compounds, identical in all respects with the 7- β -D-glucofuranosyl derivatives of zeatin and 6-BAP obtained by displacement of the 6-methylthio group. The u.v. spectra of these two compounds were consistent with their possessing an N⁶,7-disubstituted adenine chromophore¹³.

Comparison of the u.v., mass spectral (including g.c.-m.s. of the TMS and permethyl derivatives) and t.l.c. characteristics of the synthetic 7- β -D-glucofuranosides of zeatin (7) and 6-BAP (9) with the 7-glucosyl metabolites of zeatin (raphanatin)² and 6-BAP³ respectively established that these were not identical. In particular the behaviour of the metabolites on t.l.c. plates impregnated with borate buffer strongly supported the assignment of a gluco-pyranose structure to the sugar moiety in both compounds, in contradiction to the proposal by Fox. Synthesis of the 7-glucopyranosides (6) and (8) by the above route has proved unsatisfactory and alternative procedures are being examined.

A minor but highly active metabolite of 6-BAP from excised radish seedlings has been identified as 6-benzylamino-3- β -D-glucofuranosylpurine (12). The metabolite was slowly cleaved by almond β -glucosidase and showed the characteristic u.v. spectral absorptions for an N⁶,3-disubstituted adenine. 6-Benzylaminopurine was condensed with α -D-tetra-0-acetylglucopyranosyl bromide in DMF at 100° ¹⁴ and the major product separated and deacetylated to provide synthetic 6-benzylamino-3- β -D-glucofuranosylpurine (12) identical in all respects

including biological activity with the isolated metabolite. This is the first reported instance of the isolation from plant tissue of a compound with a glycosidic linkage at the 3-position of a purine ring. The biological activity of the cytokinin glucosides and their role in plant metabolism will be reported elsewhere.

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